

# Manual

## RNase-Free DNase I manual

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For Research Use Only. Not for use in diagnostic procedures.

RNase-Free DNase I is part of the Epicentre™ product line, known for its unique genomics kits, enzymes, and reagents which offer high quality and reliable performance.

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### 1. Introduction

RNase-Free DNase I is an endonuclease that efficiently hydrolyzes double- (ds) or single-stranded DNA to a mixture of short oligo- and mononucleotides. In the presence of  $Mg^{2+}$ , cleavage of each strand of a dsDNA substrate proceeds independently.<sup>1</sup> In contrast, in the presence of  $Mn^{2+}$ , the enzyme cleaves both strands of DNA at approximately the same site to generate molecules with blunt ends or 1- or 2-base overhangs<sup>1</sup> that can be blunted with T4 DNA Polymerase.

### 2. Product designations and kit components

Product	Kit size	Catalog number	Reagent description	Part numbers	Volume
RNase-Free DNase I	5,000 MBU	D9905K	DNase I, RNase-Free (1 U/ $\mu$ L)	E0013-1D4	5 mL
			10X DNase I Reaction Buffer	SS000751-D2	5.5 mL
	10,000 MBU	D9910K	DNase I, RNase-Free (1 U/ $\mu$ L)	E0013-1D4	10 mL
			10X DNase I Reaction Buffer	SS000751-D2	11 mL

### 3. Product specifications

**Storage:** Store only at  $-20\text{ }^{\circ}\text{C}$  in a freezer without a defrost cycle.

**Storage buffer:** RNase-Free DNase I is supplied in a 50% glycerol solution containing 10 mM Tris-HCl (pH 7.5), 50 mM  $\text{CaCl}_2$ , and 10 mM  $\text{MgCl}_2$ .

**DNase I 10X reaction buffer:** 100 mM Tris HCl (pH 7.5), 25 mM  $\text{MgCl}_2$ , and 5 mM  $\text{CaCl}_2$ .

**Unit definition:** One Molecular Biology Unit (MBU; also called Unit (U)) of RNase-Free DNase I produces an increase in the  $A_{260}$  of a solution of dsDNA, of 0.001 per minute at  $25\text{ }^{\circ}\text{C}$ . Functionally, 1 MBU (1 U) completely digests 1  $\mu\text{g}$  of pUC19 DNA to oligonucleotides in 10 minutes at  $37\text{ }^{\circ}\text{C}$ .

**Quality control:** RNase-Free DNase I is function-tested in two assay systems. A hyperchromicity assay is performed in a reaction containing 50  $\mu\text{g}/\text{mL}$  native calf thymus DNA, 0.1 M sodium acetate (pH 5.0), 5 mM  $\text{MgCl}_2$ , 2 mM  $\text{CaCl}_2$ , and varying amounts of enzyme. A digestion assay is performed in a reaction containing 33 mM Tris-acetate (pH 7.5), 66 mM potassium acetate, 10 mM magnesium acetate, 0.5 mM dithiothreitol, 2 mM  $\text{CaCl}_2$ , 1.0  $\mu\text{g}$  of pUC19 DNA, and varying amounts of enzyme.

**Contaminating activity assays:** RNase-Free DNase I is free of detectable RNase activities as assayed by PAGE analysis of 1  $\mu\text{g}$  of a synthetic RNA transcript following incubation with enough RNase-Free DNase I to completely digest 1 mg of DNA.

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### 4. Applications

- Elimination of the DNA template following *in vitro* transcription reactions.
- Characterisation of DNA:protein interactions by “DNase I footprinting”.<sup>1,2</sup>
- Treatment of RNA prior to RT-PCR.<sup>3</sup>
- Radiolabeling of DNA by nick translation.<sup>1,4</sup>

### 5. Example protocol

1. Dilute DNase I 10X Reaction Buffer to 1X using RNase-Free water.
2. Prepare 50 µL of a working DNase I Solution for each sample to be treated by adding 5 µL of RNase-Free DNase I to 45 µL of 1X Reaction Buffer (from Step 1).
3. Completely re-suspend 5 µg of a nucleic acid pellet in 50 µL of working DNase I solution.
4. Incubate at 37 °C for 10 minutes.

### 6. References

1. Sambrook, J. *et al.*, (1989) in: *Molecular Cloning: A Laboratory Manual (2nd ed.)*, Cold Spring Harbor Laboratory Press, New York.
2. Galas, D.J. and Schmitz, A. (1978) *Nucleic Acids Res.* **5**, 3157.
3. Kienzle, N. *et al.*, (1996) *BioTechniques* **20**, 612.
4. Rigby, P.W.J. *et al.*, (1977) *J. Mol. Biol.* **113**, 237.

### 7. Further support

If you require any further support, please do not hesitate to contact our Technical Support Team: [techsupport@lqcgroupp.com](mailto:techsupport@lqcgroupp.com)



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